

Models of grid cells and theta oscillations

ARISING FROM M. M. Yartsev, M. P. Witter & N. Ulanovsky *Nature* **479**, 103–107 (2011)

Grid cells recorded in the medial entorhinal cortex (MEC) of freely moving rodents show a markedly regular spatial firing pattern whose underlying mechanism has been the subject of intense interest. Yartsev *et al.*¹ report that the firing of grid cells in crawling bats does not show theta rhythmicity “causally disproving a major class of computational models” of grid cell firing that rely on oscillatory interference^{2–7}. However, their data may be consistent with these models, with the apparent lack of theta rhythmicity reflecting slow movement speeds and low firing rates. Thus, the conclusion of Yartsev *et al.* is not supported by their data.

In oscillatory interference models, path integration is performed by velocity-dependent variation in the frequencies of theta-band oscillations, which combine to generate the grid-cell pattern^{2–4,6,7}. In addition, learned associations to environmental sensory inputs (possibly mediated by place cells) ensure that grids are spatially stable over time and are sufficient to maintain firing in familiar environments^{2,3,8}. In rats, the majority of grid cells show theta-modulated firing^{9,10}, and the model predicts specific relationships between modulation frequency, running velocity and grid scale⁴, which have been verified in grid cells¹¹

and in putative velocity-controlled oscillatory inputs identified as interneurons within the septohippocampal circuit⁷.

Yartsev *et al.*¹ recorded the firing of grid cells from bats trained to crawl within the recording environment, a behaviour that they perform very slowly (a mean speed of 3.7 cm s^{-1} versus 17.6 cm s^{-1} in our rat data), often stopping entirely (supplementary figure 11 in ref. 1). The authors found grid cells with very low firing rates (a mean peak rate of 0.56 Hz versus 5.14 Hz in our data) and little significant theta modulation. However, matching movement speed is important for comparisons involving theta. At low speeds movement-related theta rhythmicity is strongly attenuated¹² and the need for path integration is reduced. Equally importantly, low firing rates impede detection of theta rhythmicity ($5\text{--}10 \text{ Hz}$), which requires periods containing plenty of spikes fired within tens to hundreds of milliseconds of each other (something that is absent in bat interspike interval histograms; supplementary figure 2b in ref. 1).

We examined whether differences in movement speeds and firing rates between the rat data and the bat data could explain the apparent lack of theta rhythmicity in bat grid cells. We took random samples of 25 cells from a representative data set of 85 grid cells recorded in rat MEC (Fig. 1a, bottom row), extracted periods of slow running to match bat movement speeds, and duplicated this data until it exceeded the duration of the longest bat trial (60 min). We then randomly discarded spikes to match the mean firing rates of each of the 25 published bat grid cells. From the 25 down-sampled rat cells matching each bat grid cell, we selected the one with the median theta index as representative. This process was repeated 10 times. Subjecting the 10 sets of 25 down-sampled cells to the analyses of Yartsev *et al.* produced a relative absence of theta rhythmicity (Fig. 1b, fourth row). So, if rats

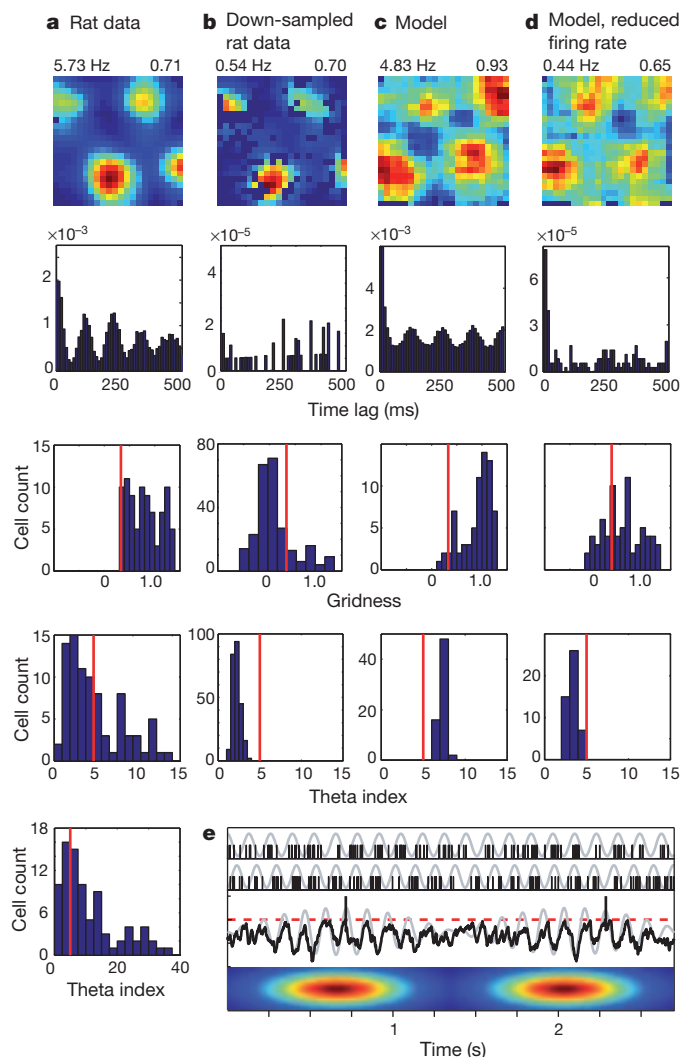


Figure 1 | Down-sampled rat grid cells and oscillatory interference reproduce bat grid-cell firing. **a, b**, The firing of grid cells in rats (**a**) resembles grid-cell firing in bats¹ if the rat data are down-sampled to match the low firing rates and slow movements of the bat data (**b**). **c, d**, The oscillatory interference model simulates theta-modulated grid cell firing in rats (**c**), and also apparently un-modulated grid-cell firing in bats when firing rates are reduced (**d**). **a–d**, Top row, example firing-rate maps (peak rate and gridness, above). Second row, example spike-train autocorrelograms. Third row, distributions of gridness scores. Fourth row, distributions of theta modulation (theta index). Grid cells have gridness > 0.33 (red line). ‘Theta-modulated cells’ have a theta index of ≥ 5 (red line). The theta index exceeded the 95th percentile for that cell’s temporally shuffled spike times for 58% of rat cells (**a**) but only for 2% of cells down-sampled to match the bat data (**b**; averaged over 10 samples of 25 cells). This rises to 14% if speed is not down-sampled, 20% if only the 25 most strongly theta-modulated rat cells are used and 72% for the 25 most strongly theta-modulated cells, if speed is unmatched. However, we do not consider this last cell population to be comparable to the bat grid cells because of the pre-selection of only the most strongly theta-modulated cells and the difference in movement speed between running rats and crawling bats. Theta index, gridness and shuffling follow ref. 1 (in which theta index is theta power divided by mean power $0\text{--}50 \text{ Hz}$), except for **a**, bottom row, which shows theta index calculated following ref. 13 (that is, theta power divided by mean power $0\text{--}125 \text{ Hz}$), giving higher values that match the proportions of theta-modulated cells in ref. 13 (which range from 62% in layer V, where most bat cells were recorded, to 90% in layer III). **e**, Schematic showing how theta-modulated inhibitory spike trains (top, black ticks) drive the grid cell’s membrane potential (middle, black trace), producing spikes when exceeding a threshold (middle, red dashed line). Spatial firing fields (bottom) are defined by constructive interference (top, grey lines show theta modulation; middle, grey line shows the resulting interference pattern), but the underlying oscillations are undetectable at low firing rates (see Methods for details).

moved as slowly as bats and their grid cells fired as infrequently, rat grid cells would show bat levels of gridness (below the higher levels seen in rats), and theta modulation would be very hard to detect.

Most importantly, to disprove the model requires knowing how much theta rhythmicity it predicts in low-firing-rate cells. Simulations (using code adapted from ref. 7) with strong theta modulation and typical firing rates for rats (Fig. 1c) also lack significant theta modulation when firing rates are reduced to bat levels (Fig. 1d, fourth row). Although spatially modulated firing is driven by interference between theta-modulated inputs, the theta rhythmicity is undetectable in low-rate spike trains (Fig. 1e).

Local field potentials and multi-unit activity were also reported in bats¹, but these reflect the physical arrangement and coherence of populations of cells, which may vary between species and are not addressed by the model (although spatially offset grids require phase-offset oscillators⁷, suggesting no overall phase preference in the model). Finally, consistent with the model, grids might be set up through oscillatory interference during the initial training of the bats to not fly out of the box (by physically blocking from above), and maintained (at lower firing rates) by learned sensory associations during subsequent slow crawling in the now highly familiar box.

Methods

The activity of 85 grid cells was recorded from superficial and deep layers of rat MEC during 20 min foraging in 1-m² arenas using standard procedures⁸. Random samples of 25 cells were speed matched by removing periods of fast running, retaining periods of ≥ 0.5 s, until the median speed was 3.7 cm s⁻¹. Speed-matched data were duplicated and concatenated to exceed the duration of the longest bat trial (60 min). Cell firing rates were down-sampled by randomly removing spikes, in turn, to match the mean firing rate of each of the 25 bat grid cells (mean rate taken as 25% of the peak rates found by Yartsev *et al.* (range of mean rates, 0.03–0.40 Hz)). Spike-train autocorrelograms combined the individual autocorrelograms from each period of slow running¹¹ and were mean-normalized to avoid low-frequency power reducing the theta index (compare with figure 4g in ref. 1). Grid cells were simulated as leaky integrate-and-fire neurons (time constant 20 ms) receiving three oscillatory inhibitory spike trains⁷ (Poisson processes with rate $50 + 30\cos(2\pi ft)$, in which frequency (f) varies around 8 Hz according to running velocity, with a peak inhibitory synaptic conductance¹⁴ of 14 pS) and a noisy persistent excitatory current sampled from $N(m, 2m)$, in which $m = 336$ nA for low firing rates and $m = 436$ nA for high rates (mean peak rates are 0.48 Hz and 5.11 Hz, respectively).

Caswell Barry^{1,2,3}, Daniel Bush^{1,2}, John O'Keefe^{4,5} & Neil Burgess^{1,2}

¹UCL Institute of Cognitive Neuroscience, 17 Queen Square, London WC1N 3AR, UK.

Yartsev *et al.* reply

REPLYING TO C. Barry, D. Bush, J. O'Keefe & N. Burgess *Nature* **488**, <http://dx.doi.org/nature11276> (2012)

Barry *et al.*¹ propose that it is impossible to detect theta rhythmicity in bat grid cells because of their slow movement velocities and low firing rates; hence, they posit that our findings² do not refute the oscillatory interference models of mammalian grid cells. To support this claim, they use a data set of rat grid cells of which only 58% were theta modulated, and constrained their analysis to periods of near immobility in the rat, a behavioural state in which theta is known to be absent³. Despite these biases, we argue that their own analysis showed that down-sampled rat cells were substantially more theta-modulated than real grid cells from bats, and we demonstrate further that the bat data have adequate statistical power to detect theta rhythmicity—if it was present in bat grid cells. Finally, Barry *et al.*

email: n.burgess@ucl.ac.uk

²University College London Institute of Neurology, Queen Square, London WC1N 3BG, UK.

³University College London Institute of Behavioural Neuroscience, 26 Bedford Way, London WC1H 0AP, UK.

⁴University College London Department of Cell and Developmental Biology, Gower Street, London WC1E 6BT, UK.

⁵Sainsbury Wellcome Centre for Neural Circuits and Behaviour, University College London, Gower Street, London WC1E 6BT, UK.

Received 18 November 2011; accepted 26 April 2012.

1. Yartsev, M. M., Witter, M. P. & Ulanovsky, N. Grid cells without theta oscillations in the entorhinal cortex of bats. *Nature* **479**, 103–107 (2011).
2. Burgess, N., Barry, C., Jeffery, K. J. & O'Keefe, J. A grid and place cell model of path integration utilizing phase precession versus theta. <http://cdn.f1000.com/posters/docs/225> (Computational Cognitive Neuroscience Conference Poster, 2005).
3. Burgess, N., Barry, C. & O'Keefe, J. An oscillatory interference model of grid cell firing. *Hippocampus* **17**, 801–812 (2007).
4. Burgess, N. Grid cells and theta as oscillatory interference: theory and predictions. *Hippocampus* **18**, 1157–1174 (2008).
5. Giocomo, L. M., Zilli, E. A., Fransen, E. & Hasselmo, M. E. Temporal frequency of subthreshold oscillations scales with entorhinal grid cell field spacing. *Science* **315**, 1719–1722 (2007).
6. Hasselmo, M. E. Grid cell mechanisms and function: contributions of entorhinal persistent spiking and phase resetting. *Hippocampus* **18**, 1213–1229 (2008).
7. Wolday, A. C., Shlifer, I. G., Bloom, M. L., Zhang, K. & Blair, H. T. Cosine directional tuning of theta cell burst frequencies: evidence for spatial coding by oscillatory interference. *J. Neurosci.* **31**, 16157–16176 (2011).
8. Barry, C., Hayman, R., Burgess, N. & Jeffery, K. J. Experience-dependent rescaling of entorhinal grids. *Nature Neurosci.* **10**, 682–684 (2007).
9. Hafting, T., Fyhn, M., Molden, S., Moser, M. B. & Moser, E. I. Microstructure of a spatial map in the entorhinal cortex. *Nature* **436**, 801–806 (2005).
10. Hafting, T., Fyhn, M., Bonnevie, T., Moser, M. B. & Moser, E. I. Hippocampus-independent phase precession in entorhinal grid cells. *Nature* **453**, 1248–1252 (2008).
11. Jeewajee, A., Barry, C., O'Keefe, J. & Burgess, N. Grid cells and theta as oscillatory interference: electrophysiological data from freely moving rats. *Hippocampus* **18**, 1175–1185 (2008).
12. O'Keefe, J. *The Hippocampus Book* (eds Andersen, P., Morris, R. G. M., Amaral, D. G., Bliss, T. V. P. & O'Keefe, J.) 475–548 (Oxford Univ. Press, 2006).
13. Boccara, C. N. *et al.* Grid cells in pre- and parasubiculum. *Nature Neurosci.* **13**, 987–994 (2010).
14. Dayan, P. & Abbott, L. F. *Theoretical Neuroscience* 180–183 (MIT Press, 2001).

Author Contributions C.B. analysed and collected experimental data; D.B. performed simulations; N.B. supervised analyses and simulations; and N.B., C.B., J.O. and D.B. contributed to writing.

Competing Financial Interests Declared none.

doi:10.1038/nature11276

focused solely on 'first generation' oscillatory interference models, ignoring our disproof of 'second generation' models. We thus uphold our original results and interpretation².

Barry *et al.* analysed a data set of 85 rat grid cells, of which only 58% were significantly theta-modulated to begin with (although oscillatory interference models require 100% of cells to be theta-modulated). The strength of theta modulation in their data was lower than in the much larger data set publically available from the Moser laboratory (a median theta index of 10.9 in Barry *et al.* compared to 14.23 in data from the Moser laboratory⁴), which may lower the detectability of theta rhythmicity after the data of Barry *et al.* is down-sampled.

Barry *et al.* proposed that bats' slow crawling velocity reduces theta rhythmicity in bat grid cells. Consequently, they selectively removed portions of the rat data, retaining short periods, down to 500 ms in duration, until a median speed of 3.7 cm s^{-1} was achieved. This procedure has several flaws.

First, although these velocities correspond to an active movement state of bats, they are equivalent to nearly complete immobility in rats^{5,6}. Barry *et al.* thus compared a state in rats in which theta is not expected³ (near immobility), to a state in bats at which theta, if existing, should be most prominent³ (active movement). Furthermore, in their model, for the constant β (which determines the velocity modulation of dendritic inputs) Barry *et al.* used a value derived exclusively from rat data⁷. However, different mammalian species would probably have different velocity dependences in dendritic inputs; for example, when modelling grid cells in cheetah versus sloth, it would not make sense to use a β value taken from rat, and the same goes for the modelling of grid cells in bats. Thus, we contend that movement speed should not be matched, neither when simulating a model nor when down-sampling data from one species to mimic another.

Second, Barry *et al.* used short portions of near-immobility data, as short as 500 ms in duration, creating very intermittent, unrealistic spike trains; this tapers down the oscillatory cycles (because they estimated 1000-ms autocorrelations using 500-ms data epochs) and could induce an unwarranted statistical bias downwards in detectability of theta rhythmicity.

Third, Barry *et al.* found that when firing rates were matched to those of bats while movement speed was left untouched, 24% of their theta-modulated grid cells retained significant theta rhythmicity (14% of 58% = 24%); substantially higher than the 4% (1/25) theta-modulated grid cells in bats². Notably, when Barry *et al.* analysed the top 51% of their theta-modulated rat grid cells (51% of the 58% modulated cells = 25 neurons)—which are the most relevant cells to consider for their model (especially given the weak theta rhythmicity in their data set)—they found that when firing rates are matched to those of bats and velocities are left untouched, the large majority (72%) of rat grid cells retained significant theta rhythmicity. Thus, down-sampled rat grid cells were markedly more oscillatory than our bat grid cells, supporting our original analysis and interpretation².

Last, Barry *et al.* considered only single-cell, first-generation oscillatory interference models of grid cells^{7–9}, which have been criticized as theoretically problematic⁹. Some of these problems have been rectified in recent second-generation versions of these models⁹, which used networks of coupled oscillators and explicitly predicted

network-level theta oscillations^{9,10}. This was contradicted by our bat data, in which brief theta oscillations occurred very rarely in the local-field potential², and multi-unit firing (reflecting network activity) never showed any theta oscillations².

In conclusion, we feel that the analysis by Barry *et al.* fails to support their main argument, namely that the statistical power of the bat data does not allow detecting theta rhythmicity. To resolve this debate, we propose to make use of a large, unbiased, publically available data set of rat grid cells, such as that on the Moser laboratory website, which would allow transparency in analysis techniques and in the baseline rat data being used. Furthermore, we expect that neural recordings from single units in flying bats, in which movement velocities and neuronal firing rates are expected to be much higher, will provide another key approach.

Michael M. Yartsev¹, Menno P. Witter² & Nachum Ulanovsky¹

¹Department of Neurobiology, Weizmann Institute of Science, Rehovot 76100, Israel.

email: nachum.ulanovsky@weizmann.ac.il

²Kavli Institute for Systems Neuroscience and Centre for the Biology of Memory, Norwegian University of Science and Technology, NO-7489 Trondheim, Norway.

1. Barry, C., Bush, D., O'Keefe, J. & Burgess, N. Models of grid cells and theta oscillations. *Nature* **488**, <http://dx.doi.org/nature11276> (2012).
2. Yartsev, M. M., Witter, M. P. & Ulanovsky, N. Grid cells without theta oscillations in the entorhinal cortex of bats. *Nature* **479**, 103–107 (2011).
3. Buzsáki, G. Theta oscillations in the hippocampus. *Neuron* **33**, 325–340 (2002).
4. Sargolini, F. *et al.* Conjunctive representation of position, direction, and velocity in entorhinal cortex. *Science* **312**, 758–762 (2006).
5. Jeewajee, A., Barry, C., O'Keefe, J. & Burgess, N. Grid cells and theta as oscillatory interference: electrophysiological data from freely moving rats. *Hippocampus* **18**, 1175–1185 (2008).
6. Wills, T. J., Cacucci, F., Burgess, N. & O'Keefe, J. Development of the hippocampal cognitive map in preweanling rats. *Science* **328**, 1573–1576 (2010).
7. Burgess, N., Barry, C. & O'Keefe, J. An oscillatory interference model of grid cell firing. *Hippocampus* **17**, 801–812 (2007).
8. Giocomo, L. M., Zilli, E. A., Fransén, E. & Hasselmo, M. E. Temporal frequency of subthreshold oscillations scales with entorhinal grid cell field spacing. *Science* **315**, 1719–1722 (2007).
9. Giocomo, L. M., Moser, M.-B. & Moser, E. I. Computational models of grid cells. *Neuron* **71**, 589–603 (2011).
10. Zilli, E. A. & Hasselmo, M. E. Coupled noisy spiking neurons as velocity-controlled oscillators in a model of grid cell spatial firing. *J. Neurosci.* **30**, 13850–13860 (2010).

Author Contributions All authors contributed to writing this reply.

doi:10.1038/nature11277